

A Model for Reversing the Cardiotoxic Effects of Doxorubicin via Fisetin in Saccharomyces cerevisiae

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Doxorubicin is a globally utilized drug for cancer treatment. This drug has unfortunate cardiotoxic side-effects through drug-induced functional loss of iron-regulatory proteins (IRP1, IRP2), enabling iron dysregulation within the cell. Doxorubicin-induced toxicity leads to downstream events, such as the overaccumulation of intracellular iron, hydroxyl radical formation through the Fenton Reaction, and caspase-9 activation, leading to cardiomyocyte apoptosis. This study aims to reverse the effects of iron overload in a eukaryotic cell model using the natural compound fisetin. Wild-type Saccharomyces cerevisiae (brewers yeast), propagated through fermentation on malt-agar, was treated with ferrous sulfate to induce iron toxicity via the Fenton Reaction. A cell viability analysis, using methylene blue as an indicator, was conducted with over 10,000 cells via a hemocytometer. Results showed that 62.8% of the ferrous sulfate-treated yeast cells died. Following fisetin treatment, in ferrous sulfate pre-treated yeast, this cell death percentage diminished to 37.4% of cells dead, 0.8% less than the control cell mortality. Statistical significance was established for the cell-death percentages across the three samples. 150 molecular docking simulations were performed and analyzed to support fisetin's ability to inhibit caspase-9, a key apoptosis regulator. Fisetin showed strong inhibition potency, with an 8.86-micromolar inhibition constant for caspase-9. A novel finding was that fisetin is bound to exosite F of caspase-9, supporting its ability to block the active site of caspase-9 and inhibit cell death. This data supports future studies investigating fisetin-mediated inhibition of doxorubicin-induced iron toxicity in multicellular organisms to make cancer treatment safer.

1. In this research project, the student directly handled, manipulated, or interacted with (check ALL that apply):

- ☐ human participants
- ☐ potentially hazardous biological agents
- ☐ vertebrate animals
- ☐ microorganisms
- ☐ rDNA
- ☐ tissue

2. I/we worked or used equipment in a regulated research institution or industrial setting (Form 1C):

YES

☒ NO

3. This project is a continuation of previous research (Form 7):

YES

☒ NO

4. My display board includes non-published photographs/visual depictions of humans (other than myself):

YES

☒ NO

5. This abstract describes only procedures performed by me/us, reflects my/our own independent research, and represents one year's work only:

☒ YES

NO

6. I/we hereby certify that the abstract and responses to the above statements are correct and properly reflect my/our own work.

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NO

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